**COMPERATIVE INVESTIGATION OF STRUCTURE OF MALE GENITAL SYSTEM OF DUCTUS DEFERENS LIGATION IN RATS**

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**ABSTRACT**

**Objective:** We investigated structural and morphological changes in pre-pubertal and pubertal rat testis and ductus deferens after unilateral vasectomy. Vasectomy is reliable and popular male contraception method, applied widely in the world.

**Methods:** 6 pre-pubertal and 6 pubertal male Wistar albino rats were divided into four groups. Unilateral ductus deferens ligation was performed on right ductus deferens of each rat. Right testis and ductus deferens tissues were evaluated as ligation group while left tissues were control groups. Two months after operation, the vas deferens and testes removed to obtain paraffin block sections and morphometric studies carried out with light microscopy**.** Hematoxylin-eosin staining and Vascular endothelial growth factor (VEGF)immunohistochemically staining were applied.

**Results:** Structural and morphological changes and decrease in spermatogenesis process have been observed at testis and ductus deferens tissues of all vasectomized rat. The seminiferous epithelium and the seminiferous tubule diameter were thickened. Degenerative changes have been observed at the epithelial structure of Ductus deferens and among muscle fibers forming the muscle of the Ductus deferens. Moreover, in pre-pubertal and pubertal vasectomized rat VEGF immunoreactivity was not observed in seminiferous tubule.

**Conclusion:** The unilateral ligation operation causes damage at ductus deferens and testis tissue in the pre-pubertal and pubertal period.

**Keyword:** Ductus deferens ligation, ductus deferens, testis, vascular endothelial growth factor, puberty, prepuberty.

**INTRODUCTION**

Vasectomy is an effective and a safe method of contraception used by men commonly all over the world (1, 2). An approximate 33 million of married women worldwide rely on their partner's vasectomy for contraception (2).

There are many controversial results about the male reproduction damage vasectomy in the literature. Many studies have been done to examine the damage and changes after vasectomy. In animal experimental studies, there was an increase in testicular degeneration in proportion to time after vasectomy operation. Compared with the findings of experimental studies on animals, no significant changes of degeneration were found in human studies. However, it was remarkable that fertility was low (3-6).

According to some researchers, vasectomy does not affect sperm production. However, since the spermatozoa produced cannot move along the duct due to occlusion, bilateral binding to the ductus deferens, sperm granulomas located along the ductus deferens and ductus epididymis are observed. In some studies on rabbits, sperm granulomas were not found. Due to the easily tearable structure of the ductus epididymis in human experiments, it is thought that the rise of pressure hinders the reflection of seminiferous tubules (5, 7-10).

In experimental studies performed, it was observed that the histological structure of testicular tissue was damaged from the first months after vasectomy and increased in proportion to time. This damage and change in Sertoli cells was less than in spermatogenic cell lines. In seminiferous tubules, vacuoles were observed in the first months, together with spermatogenic cell losses (11).

Vascular endothelial growth factor (VEGF) is an angiogenic peptide and a growth factor specific for endothelial cells. It is a principal regulator of vasculogenesis and angiogenesis (12-14). Vascular endothelial growth factor (VEGF) is known to influence testis function (15, 16). Leydig cells and other testicular cells secrete vascular endothelial cell growth factor (VEGF), whereas the receptors, VEGF-R1 and VEGF-R2 are expressed on testicular blood vessels (15, 17, 18).

The main goal of our study was the examination of structural and morphological changes in testis and ductus deferens and demonstrate alterations of VEGF immunoreactivity in rat testes after vasectomy by means of light microscopy.

**MATERIALS AND METHODS**

***Animals and experimental design***

Animals were obtained from the Animal Breeding and Experimental Research Laboratory of Gazi University. The experimental protocol was approved by the Ethical Committee of Gazi University. In this study, 6 pre-pubertal (10 days old) and 6 pubertal (60 days old) male Wistar Albino rats were used. The animals were kept at 25±2°C temperature and 32±7% humidity in a 12 h light/dark cycle. Rats were provided pelleted food and water ad libitum.

Unilateral ductus deferens ligation was performed on right ductus deferens of each rats. Right testis and ductus deferens tissues were evaluated as ligation group and left vas deferens was not ligated, left testis and ductus deferens were evaluated control group. The animals divided into 4 groups. Group 1: Pre-pubertal control group (n=6): At the beginning of the experiment vas deferens was not unilateral ligation; Group 2: Pre-pubertal vas deferens ligation group (n=6) at the beginning of the experiment unilateral vas deferens ligation performed. Group 3: Pubertal control group: (n=6) at the beginning of the experiment vas deferens was not unilateral ligation Group 4: Pubertal ligation group: (n=6). At the beginning of the experiment unilateral vas deferens ligation performed. After 2 months all the animals was sacrificed and the vas deferens and testis removed.

***Vas Deferens Ligation***

The rats were anesthetized with an intraperitoneal injection of 0.15 ml (2%) Rompun™ (xylazine) and 0.30 ml (10%) Ketasol (Ketamin). 2-3 cm midline suprapubic incision was made. The bladder and intestines were retracted and right vas deferens was identified then opened tunica vaginalis. About 10 mm tissue was resected from vas deferens and then the distal vas deferens was ligated with 5/0 silk sutures. Then, testis and other attachments were sent into scrotum. The suprapubic incision was closed with a 2-layer 3/0 vicrly. Before and after the surgical procedure, the suture area was cleaned with antiseptic solution. After two months the animals in all the groups were sacrificed and testis and vas deferens tissues removed.

***Histological and morphometric investigation***

The removed testes and vas deferens were fixed in 10% formaldehyde, then the tissues were processed the routine histological procedure and embedded in paraffin block. Tissue sections of 4–5 µm thickness were taken with a microtome. The sections stained with hematoxylin-eosin (HE). The slides were evaluated under light microscope (Leica DM4000 B, Germany). The thickness of seminiferous tubules epithelium and diameter of seminiferous tubules were measured under the light microscope with computer supported imaging system.

***Immunohistochemical Examination***

For immunohistochemical analyses 4-5 µm-thick sections were taken with microtome (Leica MR 2145). VEGF (Ab-1) (Cat: RB- 222-P1, Lot:222p810C, Neomarkers, Fremont, CA) rabbit polyclonal antibody was used for the immunohistochemical studies to investigate vascularization in testes. After deparaffinization, rehydration and leaving in distilled water for 5 min, the samples were incubated in 1 M EDTA buffer (pH: 6.0) ((pH: 6. 0) (Cat: AP- 9004- 500, Lot: AX201103, Lab Vision, Fremont, USA). Then, the sections were incubated in 3 % H2O2 (Cat: TA-125- hp, Lot: HP18180, Lab Vision, Fremont, USA) for 15 min to inhibit endogenous peroxidase activity and after H2O2 treatedthe testes sections were washed in phosphate buffer solution (PBS). UltraV block (Cat: TA-125-UB, Lot: AUB110414, Lab Vision, Fremont, USA) was applied to prevent non-specific binding and following the blocking, the sections were incubated with primary antibody VEGF (1:100 dilution) for 60 min. Then, the sections were washed in PBS and incubated with biotinylated secondary antibody (Cat: TR-125-BN, Lot: RBN100924, Lab Vision, Fremont, USA) for 20 min. After washing with PBS, the sections were conjugated with streptavidin peroxidase complex (Cat: TS-125- HR, Lot: SHR110303, Lab Vision, Fremont, USA) for 20 min. Finally, sections were incubated with AEC (3- amino-9-ethylcarbazole) (Cat: TA-125-SA, Lot: ASA110429, Lab Vision, Fremont, USA Mayer’s hematoxylin (Cat: TA-125-MH, Lot: AMH70809, Lab Vision, Fremont, USA) was used for background staining. The sections with Ultramount medium (Cat: TA-125-UG, Lot: VM13518, Lab Vision, Fremont, USA) were ready for microscopic examination. Immunohistochemical analyses of VEGF were studied in testes paraffin sections via imaging system (Leica Q Vin 3).

**Statistical Analysis**

Statistical analyses of data were performed with the Statistical Package for Social Sciences (SPSS) v.11 software for Windows. Non-parametric data was analyzed Mann-Whitney U test. P-values < 0.05 were considered statistically significant.

**RESULTS**

***HE Staining***

According to our histological results we observed that the ovale or irregular seminiferous tubules and immature interstitial area in testes tissue of pre-pubertal control group (Figure 1.A, 1.B). Leydig cells and blood vessels were clearly distinguished in interstitium. Seminiferous tubules including that more immature differentiation stage spermatogenic series such as spermatogonia, spermatocytes. Sertoli cells have been observed along the basement membrane.

When we evaluated the testes of pre-pubertal vas deferens ligation group, degeneration and irregular boundaries of seminiferous tubules were observed. Irregularities in cells of the spermatogenic series were seen. Sertoli cells were not clearly distinguished in the basal compartment. Degeneration and necrosis of interstititum and fewer Leydig cells have been observed (Figure 1.C, 1.D).

The normal histological structure of the testes was observed in the cells of spermatogonial series, the Sertoli and Leydig cells in the pubertal control group. The interstitium is more regular appearance compared with the control pre-pubertal control group. (Figure 1.E, 1.F)

In the testes section of pubertal vas deferens ligation group, degeneration in seminiferous tubules and significant loss of the developing spermatogenic cell series were observed. The composition of interstitium was not clearly distinguished (Figure 1.G, 1.H).

The histological analysis of vas deferens in pre-pubertal control group clearly revealed that pseudostratified columnar epithelium and lamina propria appeared normal. It was noted that the muscular layer consisted of muscle fibers that were still in one direction compared to the pubertal control group. Tunica adventitia had numerous nerves and blood vessels, but that the connective tissue fibers and cells were not clearly observed (Figure 2.A).

Microscopic examination of vas deferens in the vasectomized pre-pubertal rats was showed that luminal epithelium and underlying tissues were separated from each other and tissue integrity impaired (Figure 2.B). There were degenerative changes between pseudostratified columnar epithelial cells stereo cilia of which was different lengths and muscular layer (Figure 2.B).

Pseudostratified columnar epithelial cells stereo cilia of which was different lengths

The vas deferens of pubertal rat group showed that normal histologic structure. Luminal epithelium, lamina propria, muscular layer and tunica adventitia were normal morphology (Figure 2.C).

In the vasectomized pubertal rats, irregular morphology for stereosilium, muscle fiber of muscular layer and thinned lamina propria were seen comparison to the control sections of pubertal rat. Connective tissue content was altered and a decrease in the structural integrity was observed in tunica adventitia layer compared to control group (Figure 2.D).

***Immunohistochemically evaluation***

*VEGF Staining*

In the pre-pubertal and pubertal control group, VEGF immunoreactivity was observed for the spermatogenic cells in the seminiferous tubules and in the endothelial cells of the blood vessels (Figure 3.A). According to our immunohistochemical findings, we found no VEGF positive staining in the spermatogenic series and endothelium of blood vessel in vasectomized pre-pubertal and pubertal rats (Figure 3.B). VEGF immunostaining was observed strongly for spermatogenic series and Sertoli cells in compared to ductus deferens, pubertal group (Figure 3.C). VEGF immunoreactivity was not observed for the sections of testis in the ligation group of pubertal rat (Figure 3.D).

***Statistical Analysis***

*Evaluation of Diameter and thickness of the seminiferous tubules*

Diameter and thickness of the seminiferous tubules were measured in six cross-sections from each experimental group. Mean ± Standard deviation, median and minimum /maximum values were displayed in table. The diameter and epithelium thickness of seminiferous tubule of rats from pubertal control group was significantly higher than pubertal ligation group (p=0.000). We found that the diameter and epithelium thickness of seminiferous tubule of pre-pubertal control group rats, were significantly higher compare to of those of ligation group. (p=0,000)

**DISCUSSION**

Vas deferens ligation (=Vasectomy), is a safe procedure in male contraception and use widely in the world (5). The experimental data is rather controversial and there is no general agreement about the effect of vasectomy on testicular and vas deferens histology.

In order to investigate the effects and damage of vasectomy, degenerative changes in the testicular tissue have also been observed following vasectomy in several species including human (19, 20), guinea pig (21), rabbit (22), dog (23) and monkey (24). Moreover, the histological structure of testicular tissue was damaged after vasectomy and increased in proportion to time (25). However; after vasectomy operation, some recovery at different times was also shown at humans (at 2 years) (19) and rats (at 1 to 3 months) (26).

In our study, we examined that the effects of unilateral vas deferens ligation on histological alterations in the structure of the testis and vas deferens and VEGF immuno-expression changes in the testis of the pre-pubertal and pubertal rat.

We found that ductus deferens ligation caused some histological changes in the testis, decreased seminiferous tubule size, and that the development of the spermatogenic cell line was affected at an advanced stage in present study.

Process of the vasectomy suddenly prevents the flow of sperm and seminal fluid, resulting in a sudden increase in pressure in the seminiferous tubules. Such degeneration takes places through pressure damaging the seminiferous epithelium (27). These changes may vary depending on the area where the obstruction is located (28, 29).

Vas deferens ligation also has negative effects on spermatogenesis, which is the process that including germ cell proliferation and differentiation. Spermatogenic degeneration after vas deferens ligation has been reported in many studies (6, 7, 19, 22, 30).

Flickinger et al. previously described the vasectomy was not having a negative effect on the testes and rat testis remained a normal structure after vasectomy in pubertal and pre-pubertal period (31, 32).

However, as a result of some studies, it has been determined that the spermatogenesis process has deteriorated after vasectomy and the histological structure of the testis has been damaged. Neaves et al. stated that reduction in tubule diameter, depletion of advanced germinal cells and fusion of spermatids in vasectomized rats after 3 months’ surgery (33).

In the light of this information, the studies have focused especially on vas deferens obstruction (27, 31-37). In this study, testicular degeneration was associated with decreasing the diameter of the seminiferous tubule and thickness epithelium at both pre-pubertal and pubertal period. Then, it was noticed that the primary spermatocytes, secondary spermatocytes and spermatids series following the spermatogonia on the basal membrane could not be distinguished in a regular and clear form in the vas deferens ligation group. Our study agreed with previous literature as much as the diameter of seminiferous tubules were altered and decreased. In addition, we observed that testicular atrophy in vas deferens ligation group. Degeneration at pre-pubertal groups was much more than pubertal groups. The testes continue to produce sperm after vasectomy but, does not leave the body. As a result of pressure in epididymis, sperm granulomas are observed in the epididymis and vas deferens. McDonald et al, sperm granuloma develops following vasectomy (8). At other studies, sperm granulomas may be composed at the vasectomy site in some rats in 1 week-3 months after vasectomy surgery operation (3, 4, 38). Conversely, in our study sperm granulomas were not seen in testes and vas deferens. Grupta et al have been investigated that effects of vasectomy in men and observed after 1month vasectomy comprehensive degeneration of the germinal epithelium and some intertubular fibrosis (19). In this study, it was also determined that degeneration of intersitisium and loss of connective tissue cells in vasectomized pubertal and pre-pubertal rats.

Mc. Vicar et al showed that the least affected cells from the vasectomy were Sertoli cells, whereas the most affected were spermatogenic cells (11, 15). Furthermore, Aydos at all reported that one month after vasectomy, vacuoles and loss of spermatogenic cells were seen in the seminiferous tubules, but the Leydig cells did not show any changes (3). At other experiment, Sarrat et al have noticed influence of vasectomy on rats and dogs after four months following ligation operation. They observed that changes in the thickening of the basement membrane, atrophic the seminiferous tubules, hypertrophy of the interlobular interstitial tissue, while they did not see an increase in the number of Leydig cells (23). Similar to these researches, there were no any changes at Sertoli cell and Leydig cell in our study.

Vascular endothelial growth factor (VEGF) is a major regulator of physiological and pathological angiogenesis (12-14). Nalbandian et al. investigated that VEGF is expressed in Sertoli cells, and VEGFR-2 and VEGFR-1 receptors are expressed in spermatogonial stem cells and advanced germ cell types, respectively. The authors have concluded that VEGF may play a potential regulatory role in the spermatogenesis process with VEGFR-1 and VEGFR-2 (39). In another study demonstrated that VEGF has been implicated in both angiogenesis and seminiferous cord formation (40).

In our histochemical findings, VEGF immunoreactivity was observed for the spermatogenic cells in the seminiferous tubules and in the endothelial cells of the blood vessels in the pre-pubertal and pubertal control group. In contrast, we found that VEGF immunopositivity were not detected in the spermatogenic series and endothelium of blood vessel in vasectomized pre- and pubertal all rats.

VEGF immunostaining was observed strongly for spermatogenic series and Sertoli cells in compared to vas deferens ligation pubertal group while VEGF immunoreactivity was not observed for the sections of testis in the ligation group of pubertal rat. Many studies have previously emphasized, there is a significant reduction in the number of spermatogenic cells and interstitial fibrosis developed among seminiferous tubules after vasectomy or vas deferens ligation. Therefore, it is possible to explain the absence of VEGF immunoreactivity because of the damage of the cells on the testes in vasectomy group according to the control group.

In summary, it is possible to propose VEGF as an intratesticular regulatory factor being crucial for the maintenance of normal physiological function of the testis.

In conclusion, we showed that the bilateral vasectomy operation may have effects on male infertility due to changes in testes and ductus deferens at pre-pubertal and pubertal period. However, it was considered necessary to support it with another methods and clinical investigations.

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